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## Stereoselective synthesis of the C33-C44 fragment of palau'amide

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## Abstract

An efficient stereoselective synthesis of the C33–C44 fragment of palau'amide is described using a Sharpless asymmetric epoxidation, a regioselective nucleophilic ring opening of the epoxide, a Grignard reaction and a Luche stereoselective reduction of a keto compound as the key steps.

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In 2003, Moore and co-workers reported the isolation, structure elucidation, and biological activity (cytotoxicity to KB cells,  $IC_{50} = 13 \text{ nM}$ ) of palau'amide (1), an architecturally novel cyclic depsipeptide from the bioassay-guided fractionation of the extract species of Lyngbya from Palau.<sup>1</sup> Key structural elements include a five amino acid backbone fused together in a macrocycle and a novel polyketide chain incorporated into the molecule. The molecular architecture of the polyketide comprises three contiguous chiral centres, a 1,3-syn diol flanking an anti methyl group, a terminal alkyne and an  $\alpha,\beta$ -unsaturated acid (Fig. 1). Potent biological activity coupled with unique structural features and limited availability prompted us to explore the synthesis of palau'amide (1). The first synthesis of palau'amide was reported by Ma et al.<sup>2</sup> The polyketide chain was synthesized following Oppolzer's protocol and utilized a vinylogous Mukaiyama aldol reaction as a key reaction. The 'anti' aldolization strategy to build the C38 and C39 stereocentres was not successful. 'svn' Aldolization and Mitsunobu inversion were required to obtain the required stereochemistry. However, the data for the synthetic palau'amide (1) did not match the reported values. A highly stereoselective and practical approach to construct the three contiguous chiral centres present in the



Fig. 1. Structure of palau'amide 1.

C33–C44 fragment was therefore required to establish conclusively the structure of the natural product and to furnish additional analogues for testing.

Our retrosynthetic analysis of fragment 2 (Scheme 1) revealed that it could be synthesized from intermediate 3 which, in turn, could be prepared from fragment 4 following oxidation and a Grignard reaction with pentynyl-magnesium bromide. Intermediate 4 was envisaged to be obtained by regioselective opening of epoxide 5, which in turn could be prepared from commercially available 1,3-propane diol following a known protocol.

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Scheme 1. Retrosynthetic analysis.

Chiral epoxide **5** was elaborated from 1,3-propane diol via a five-step sequence following the literature methods.<sup>3</sup> A key reaction was the Sharpless asymmetric epoxidation with titanium tetraisopropoxide and *t*-butylhydroperoxide in the presence of (–)-DIPT. The enantiomeric excess of epoxide **5**<sup>4</sup> was determined by HPLC analysis<sup>5</sup> and was found to be greater than 97%. Regioselective nucleophilic opening of **5** with Me<sub>2</sub>CuCNLi<sub>2</sub><sup>6</sup> afforded a mixture of the desired 1,3-diol **6** and the corresponding 1,2-diol in a ratio of 8:1. The minor product was easily removed by treatment with NaIO<sub>4</sub> (Scheme 2). Chemoselective benzoylation of the primary hydroxyl group of **6**, followed by silylation of the secondary hydroxyl group using TBSOTf<sup>7</sup> and 2,6-lutidine in dichloromethane and subsequent hydrolysis of the benzoyl ester, afforded alcohol **4**<sup>8</sup> in excellent yield.

Oxidation of the primary hydroxyl group of **4** under Swern oxidation conditions<sup>9</sup> gave an aldehyde, which on treatment with trimethylsilyl protected pentynylmagnesium bromide in THF at 0 °C furnished diastereomeric alcohols 7 and 8 in a ratio of 1:1.5. The stereochemistry at the newly created stereogenic centers in  $7^{10}$  and  $8^{11}$  was assumed to be *R* and *S* resulting from 1,3-asymmetric induction based



Scheme 2. Synthesis of 7 and 8.

on a model proposed by Evans and co-workers.<sup>12</sup> This assumption was further confirmed using the method reported by Rychnovsky and co-workers.<sup>13</sup> Accordingly, cleavage of the silyl ether in 7 and 8 produced diols, which were protected as 1,3-diol acetonides 9 and 10 by treatment with dimethoxypropane in the presence of a catalytic amount of *p*-TSA. Since the C(2)-acetals in 9 and 10 showed <sup>13</sup>C chemical shifts at  $\delta$  97.94 and 100.62 ppm and the differences between the two <sup>13</sup>C methyl signals in 9 and 10 were 10.12 and 0.09 ppm, respectively, the *syn* and *anti* juxtaposition of the acetonide moieties was confirmed (Scheme 2).

To obtain the desired alcohol **13** exclusively, the mixture of alcohols **7** and **8** was treated with  $K_2CO_3$  and MeOH to effect TMS deprotection. The resulting secondary hydroxyl group was oxidized by the Swern oxidation protocol to give keto compound **12**, which was reduced by Luche's<sup>14</sup> procedure using NaBH<sub>4</sub> and CeCl<sub>3</sub> in MeOH at -100 °C to

afford 13 as a single isomer (Scheme 3). The secondary alcohol moiety of 13 was protected as its methoxymethyl ether using MOMCl and DIPEA to afford 14 in 82% yield. The *p*-methoxybenzylether of 14 was cleaved using DDQ in CH<sub>2</sub>Cl<sub>2</sub> to furnish alcohol 3 in 91% yield. Alcohol 3 was oxidized to the aldehyde with IBX; further treatment with the Wittig reagent allyloxycarbonylethylenetriphenylphosphorane in refluxing THF gave only *E*-isomer 15 as a single



Scheme 3. Synthesis of 2.

product in an 80% yield over two steps. Finally, deprotection of the *tert*-butyldimethylsilyl group with tetrabutylammonium fluoride afforded the C33-C44 core fragment 2 with the required stereocenters in 86% yield. The <sup>1</sup>H, <sup>13</sup>C NMR and elemental analysis of fragment  $2^{15}$  were in good agreement with the assigned structure. For example, the <sup>1</sup>H NMR spectrum revealed signals due to the olefinic protons at  $\delta$  6.94 (m, 1H), 5.97 (m, 1H) and 5.39–5.20 (m, 2H), which were characteristic of an  $\alpha$ ,  $\beta$ -unsaturated ester and a terminal olefin group. A signal due to the alkyne proton appeared at  $\delta$  1.97 (t, J = 2.6 Hz), the methyl group attached to the double bond carbon appeared as a singlet at  $\delta$  1.89 and the methyl group present between the two hydroxyl groups appeared as a doublet at  $\delta$  0.88 (d, J = 6.9 Hz). In the <sup>13</sup>C NMR spectrum, the corresponding olefinic carbons appeared at 138.9, 132.4, 129.5 and 117.7 ppm, respectively. The carbonyl carbon of the ester group resonated at  $\delta$  167.5 ppm.

In conclusion, a practical and stereoselective synthesis of the C33–C44 fragment of palau'amide has been achieved using a Sharpless asymmetric epoxidation, a regioselective nucleophilic epoxide opening, a Grignard reaction, a Luche stereoselective reduction and an *E*-selective Wittig reaction. Further investigations towards the total synthesis of palau'amide are in progress.

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- 4. Analytical and spectral data of **5**:  $[\alpha]_D^{25} 20.34$  (c 1.7, CHCl<sub>3</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  7.24 (d, J = 8.7 Hz, 2H), 6.86 (d, J = 8.7 Hz, 2H), 4.44 (s, 2H), 3.87 (dd, J = 2.6, 12.6 Hz, 1H), 3.80 (s, 3H), 3.64–3.53 (m, 3H), 3.08 (m, 1H), 2.95 (m, 1H), 2.00–1.74 (m, 3H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  158.9, 129.9, 128.9, 113.5, 72.3, 66.2, 61.5, 58.3, 54.8, 53.4, 31.7. Anal. Calcd for C<sub>13</sub>H<sub>18</sub>O<sub>4</sub>: C, 65.50; H, 7.56. Found: C, 65.35; H, 7.48.
- 5. Enantiomeric purity of the product formed was verified by HPLC analysis. HPLC conditions: column, CHIRALCEL [OJ-H (5  $\mu$ m spherical) 250 × 4.6 mm]; mobile phase, IPA-petroleum ether (1:9); flow rate, 0.5 mL/min; UV detection at 220 nm.
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- 8. Analytical and spectral data of 4:  $[z]_D^{25} 1.32$  (c 1.7, CHCl<sub>3</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  7.24 (d, J = 8.7 Hz, 2H), 6.87 (d, J = 8.6 Hz, 2H), 4.41 (ABq, J = 11.6, 13.1 Hz, 2H), 3.89 (q, J = 5.9 Hz, 1H), 3.81 (s, 3H), 3.69 (dd, J = 3.9, 10.9 Hz, 1H), 3.50 (t, J = 6.5 Hz, 3H), 2.69 (br s, 1H), 1.88–1.74 (m, 3H), 0.97 (d, J = 7.1 Hz, 3H), 0.90 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  159.1, 130.3,

129.2, 113.7, 73.6, 72.7, 66.4, 65.1, 55.1, 39.1, 34.2, 25.8, 18.0, 13.7, -4.4, -4.6. Anal. Calcd for  $\rm C_{20}H_{36}O_4Si:$  C, 65.21; H, 9.78. Found: C, 65.24; H, 9.69.

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- 10. Analytical and spectral data of 7:  $[\alpha]_{D}^{25} + 3.05 (c 1.3, CH_2Cl_2); {}^{1}H NMR$ (200 MHz, CDCl<sub>3</sub>):  $\delta$  7.22 (d, J = 8.7 Hz, 2H), 6.84 (d, J = 8.7 Hz, 2H), 4.40 (s, 2H), 4.02 (m, 1H), 3.78 (s, 3H), 3.53–3.44 (m, 3H), 2.33 (dt, J = 2.02, 7.07 Hz, 2H), 2.20 (m, 1H), 1.94–1.44 (m, 7H), 0.86 (s, 9H), 0.82 (d, J = 6.9 Hz, 3H), 0.12 (s, 9H), 0.04 (s, 3H), 0.02 (s, 3H); {}^{13}C NMR (50 MHz, CDCl\_3):  $\delta$  159.1, 130.2, 129.3, 113.7, 107.5, 84.7, 72.8, 72.7, 71.5 67.0, 55.1, 44.2, 33.6, 33.3, 25.8, 18.0, 16.2, 11.6, 0.18, -4.4, -4.5. Anal. Calcd for C<sub>28</sub>H<sub>50</sub>Si<sub>2</sub>O<sub>4</sub>: C, 66.40; H, 9.88. Found: C, 66.32; H, 10.14.
- Analytical and spectral data of 8: [α]<sub>D</sub><sup>25</sup> −2.85 (c 0.7, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>): δ 7.22 (d, J = 8.5 Hz, 2H), 6.86 (d, J = 8.5 Hz, 2H), 4.39 (ABq, J = 11.6, 19.4 Hz, 2H), 4.12 (m, 1H), 3.95 (dt, J = 2.4, 6.8 Hz, 1H), 3.80 (s, 3H), 3.54 (m, 1H), 3.46–3.38 (m, 2H),

2.26 (t, J = 6.9 Hz, 2H), 1.94 (q, J = 6.7 Hz, 2H), 1.81–1.63 (m, 2H), 1.55–1.25 (m, 3H), 0.99 (d, J = 7.1 Hz, 3H), 0.88 (s, 9H), 0.13 (s, 9H), 0.09 (s, 6H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  159.1, 130.2, 129.2, 113.7, 107.1, 84.5, 76.0, 72.6, 69.1, 66.2, 55.1, 38.6, 35.0, 33.5, 25.8, 17.9, 16.6, 11.3, 0.2, -4.4, -4.7. Anal. Calcd for C<sub>28</sub>H<sub>50</sub>Si<sub>2</sub>O<sub>4</sub>: C, 66.40; H, 9.88. Found: C, 66.51; H, 10.02.

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- 15. Analytical and spectral data of **2**:  $[\alpha]_D^{25} 16.71$  (c 1.0, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  6.94 (m, 1H), 5.97 (m, 1H), 5.39–5.20 (m, 2H), 4.68–4.66 (m, 3H), 4.64 (t, J = 1.3 Hz, 1H), 3.81–3.63 (m, 2H), 3.40 (s, 3H), 2.78 (br s, 1H), 2.48–2.16 (m, 4H), 1.97 (t, J = 2.6 Hz, 1H), 1.89 (s, 3H), 1.76–1.55 (m, 5H), 0.88 (d, J = 6.9 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>);  $\delta$  167.5, 138.9, 132.4, 129.5, 117.7, 95.7, 84.1, 79.7, 73.2, 68.5, 65.1, 55.9, 41.5, 34.1, 29.3, 23.6, 18.3, 12.6, 12.0. Anal. Calcd for C<sub>19</sub>H<sub>30</sub>O<sub>5</sub>: C, 67.40; H, 8.87. Found: C, 67.54; H, 8.69.